

Antiseptic Effect of Betel Quid Extract on Lip Mucosal Wound of male Wistar (*Rattus norvegicus*) Rats

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Abstract

Antiseptic is commonly used in dentistry practices, but some antiseptics have side effects such as allergy. Therefore, natural substances are developed as substitute, i.e betel quid which consisted of betel leaf, areca nut, gambier, and mineral slaked lime.

The aim of this study was to determine antiseptic effect of betel quid extract on lip mucosal wound of male Wistar (*Rattus norvegicus*) rats.

This study was true experimental in vivo. Thirty rats were divided into five groups; three groups were treated with betel quid extract ointments at concentration of 5%, 10% and 20%, one group was positive control (hyaluronic acid) and last group was negative control (placebo). One-mm diameter of wound was made on lower lip mucosa of rats with cylinder diamond bur. All wounds were swabbed before and after treatment, and the number of bacterial colonies grown in agar plates were counted.

The results showed that there was significant decrease of bacterial colony numbers for respective groups, except for negative control ($p < 0.05$). Ointment containing 20% betel quid extract had antiseptic effect similar to 0.2% hyaluronic acid.

It can be concluded that betel quid extract had antiseptic effect on lip mucosal wound.

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Introduction

Antiseptic is chemical compound used to destroy the structure or to inhibit the growth of microorganisms on living tissue such as skin, mucosa and open wounds.¹ In dentistry practice, antiseptic is widely used before the implementation of teeth extraction procedures, biopsies, scaling and other procedures that have potential to injure soft tissue in oral mucosa.² The antiseptic addition has objective to provide aseptic condition in injured mucosa in order to prevent bacterial entrance into circulation and to control infection that can inhibit wound healing process.^{1,3}

One example of antiseptic substance that frequently used on oral mucosal wound is *hyaluronic acid gel* at concentration of 0.2 %.⁴ However, some studies showed that the use of topical medications of *hyaluronic acid* had side effect such as allergic reaction.⁵ Therefore, natural topical medications that has a role as antibacterial substance is required to help wound healing process. One of natural substances which contain antibacterial substance is betel quid.

Chewing betel quid is prevalent in some part of Asian countries, such as Indonesia, Malaysia, India and Taiwan.^{6,7} Traditionally, most people believed that betel quid had several medical effects such as anti-inflammation, antiseptic, anti ulcer, antimicrobial and antivirus.⁸ Betel quid, commonly chewed in Indonesia, is a mixture of betel leaf (*Piper betle* Linn), areca nut (*Areca catechu*), gambier (*Uncaria gambir* Roxb.) and mineral slaked lime (calcium hydroxide). Each components of this mixture contain

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antibacterial substances as follows: 1.3% and 4.5% of tannin and essential oil for betel leaf, 0.6% of tannin for areca nut, as well as 2-4% of quercetin and 2.7-33 % catechin for gambier.⁹⁻¹¹ Mineral slaked lime/betel lime (calcium hydroxide) has good antibacterial characteristics through division mechanism of calcium and hydroxyl ions into enzymatic reaction on bacteria that will inhibit DNA replication process.^{12,13}

Hoque et al. had reported that ethanol extract from betel leaf at the dose of 10 g/ml was capable to inhibit the growth of microorganisms on food contaminated by pathogens such as *Vibrio cholerae*, *Escherichia coli*, *Shigella dysenteriae* and *Staphylococcus aureus*.¹⁴ Rahman et al. stated that ethanol extract from areca nut seed at the dose of 20 mg/l had ability to inhibit the growth of microorganisms of *Bacillus subtilis* and *Staphylococcus aureus*.¹⁵ Katu et al. conducted antibacterial test on gambier extract and showed that ethanol extract from gambier at concentration of 1 % was effective in inhibiting the growth of bacteria of *Enterococcus faecalis*.¹⁶ Mohammadi et al. exhibited the antimicrobial activity of calcium hydroxide in root canal bacteria.¹⁷

Antibacterial and antiseptic effect of each constituents in betel quid has been done in *in-vitro* study. Study about the effect of the mixture of all components consisted in betel quid in *in vivo* would be important to be done. The objective of this study was to determine the antiseptic effect of betel quid extract on lower lip mucosal wound of male Wistar (*Rattus norvegicus*) rats.

Materials and methods

This study was true experimental *in vivo* with pretest-posttest only control group design. The study was conducted at Animal House of Medical Faculty of Sriwijaya University Palembang to do the research and at Microbiology Laboratory of Medical Faculty of Sriwijaya University Palembang to determine the number of bacterial colonies. It had been approved by Research Ethical Commission of Mohammad Hoesin General Hospital (RSMH) Palembang and Medical Faculty of Sriwijaya University with ethical certificate No. 390/kepkrsmhfkunsri/2017.

Wistar (*Rattus norvegicus* L.) rats were obtained from Pharmacy School of Bandung

Institute of Technology (SF-ITB), with certificate no. 524.3/3691-Dispangtan/2017. Inclusion criterias were male white rats of Wistar (*Rattus norvegicus* L.) strain, 8-12 weeks old, body weight of 150-200 grams and in healthy condition without defect. Rats that had been used in the previous study were excluded. Betel quid components, including betel leaf (*Piper betle* L.), areca nut (*Areca catechu* L.), gambier (*Uncaria gambir* Roxb.) and mineral slaked lime (calcium hydroxide) were collected from Babatoman Village, Sekayu Subdistrict, Musi Banyuasin District, South Sumatra Province, Indonesia. All the constituents and material were identified and authenticated by Faculty of Agriculture, Sriwijaya University, Indonesia.

Preparation of Animal

Thirty male Wistar rats were divided into 5 groups, Group A was treated with 5% betel quid extract ointment, Group B was treated with 10% betel quid extract ointment, Group C was treated with 5% betel quid extract ointment, Group D was positive control, treated with 0.2% hyaluronic acid ointment (purchased from Ricefarma Pharm.Co, Surabaya, Indonesia), and Group E was negative control, treated with placebo ointment.

Samples were acclimated for 8 days at room temperature of 20-25°C.¹⁸ During acclimatization process, rats were fed with standard pellet diet and water *ad libitum*.

Preparation of betel quid extract

Betel quid was extracted by using soxhletation method. Betel quid components consisting of 8 g of betel leaf, 3.5 g of areca nut, 2.5 g of gambier and 2 g of betel lime were mixed. Soxhlet system was filled with 96% ethanol solvent at ratio of 1:4 and heated at temperature of 50° C for five hours. The crude extract was vaporized to dryness by using rotary evaporator (IKA RV10, Staufen, Germany) for 2 days.

Preparation of ointment

Preparations of betel quid extract ointment for respective treatment groups was 5 g and the dose given to experimental rats was 50 mg. The basic materials for ointment were lipid based material consisted of 15% *adepts lanae*

and 85% *vaselin album*. Each concentrations of 5%, 10% and 20% was made as following formula :

$$M1.V1=M2.V2$$

The hot mortar and pestle was taken out from oven (Cole-Palmer Ltd, UK) and adeps lane was poured into mortar and stirred by using pestle until melt. Subsequently, vaseline album was poured into mortar and was stirred by using pestle at constant velocity until homogenous and form ointment base. Betel quid extract was added according to formulation of each groups and was stirred until homogenous. Ointment extract was flattened on surface of glass base for subsequent homogeneity test. An ointment was considered homogenous if particles mixture was evenly distributed. Homogenous ointment preparations extract was put into ointment pot and was labeled according to treatments.

Induction of mucosal wound

Mucosal wound induction on lower lip of rats was done by using 1-mm diameter of cylinder diamond bur (Microdont, USA). Prior to wound induction, animals were anesthetized with 0.2 mL ketamine by i.m. injection. The lower lip of rats was withdrawn by using tweezer (Fisher brand™, Thermofisher Co, UK) and was swabbed with aquadest-wetted sterile cotton. Mucosal wound was made by using cylinder diamond bur at depth of about 1 mm in accordance to bur diameter used in this process. Blood was cleaned with aquadest-wetted sterile cotton and dried.

Topical wound application

Before giving treatment, each wound was swabbed by using sterile cotton swab with back-and-forth motion for two seconds. Pre-test specimens were placed into sterile transport container and sent to the microbiology laboratory. The treatments were conducted according to respective groups by evenly rub cotton buds on injury part, left it for one minute and followed by conducting re-swab as the previous stage. Post-test specimens were placed into sterile transport container and sent to microbiology laboratory.

Bacterial Culture

The specimens were put into 10 mL test tube containing 1 mL of NaCl, homogenized and covered with sterile cotton. The suspension was diluted in ratio 1:10, and then the tube was rotated for 30 seconds. One mL homogenate suspension was taken from the tube by using micropipette and then put into agar media, flattened by using sterile loop and labeled according to treatments of each groups. All agar plates were incubated within incubator (ThermoFisher Co, UK) at temperature of 37°C for 24 hours. The number of bacterial colony was counted.

Antiseptic assay

The number of bacterial colonies growth in agar plates was counted by using automatic colony counter camera (Colonizer, Sunil Laxman Lab, India). Plate was put on previously prepared pad. A light transmission was used to help lighting. Plate containing bacterial colonies was photographed and was awaited for about 30 seconds. The results were shown on screen display which was connected to camera. Data were recorded and analyzed.

Statistical Analysis

The analysis was performed with Statistical Package for Social Sciences version 22 (IBM® inc.pvt ltd, US) and Microsoft Excel (Microsoft inc®, Redmond). Data were tested with Levene's test to know the homogeneity of samples and normality test of Shapiro Wilk to know the distribution of samples. P value of > 0.05 was considered that data were homogen and normal. Paired t-test was used to compare the changes between "before and after" study. One-way Anova was used for significance of difference in all groups. Post-hoc Tukey's test was continued to evaluate the effect of antiseptic among the groups and to compare the effect with positive control group. Statistically significant differences were indicated by p value of <0.05.

Results

The number of bacterial colonies on mucosal wound was evaluated. Normality test using Saphiro Wilk and homogeneity test using

Levene's test showed p value of >0.05. It meant that all data were normally distributed and homogen. All groups showed the reduction of bacterial colonies, except group E which treated with pachebo oinment. Table 1 described that betel quid extracts in all concentrations were effective in decreasing bacterial colonies significantly (p value<0.05) in which hyaluronic acid showed the best results followed by betel quid extract of 20%, 10% and 5% concentration.

One-way Anova test was performed, and the results was p=0.00. It meant that there was a significant different difference in reducing the number of bacterial colonies between and within groups after treatment. Analysis was continued to Post-hoc Tukey's test (Tabel 2).

Group	Means ± SD		P value
	Before	After	
5% betel quid extract	115.83 ± 9.99	101.67 ± 8.94	0.00*
10% betel quid extract	119.67 ± 16.11	78.00 ± 15.53	0.00*
20% betel quid extract	121.00 ± 14.34	62.50 ± 13.94	0.00*
Positive control (hyaluronic acid)	125.17 ± 11.00	49.33 ± 5.99	0.00*
Negative control (placebo)	120.17 ± 5.57	152.33 ± 12.47	0.00*

Table 1. The number of bacterial colonies before and after treatment. * Significant level, Paired t test, p<0.05.

Groups/P value	5% betel quid extract	10% betel quid extract	20% betel quid extract	Positive control	Negative control
5% betel quid extract		0.02*	0.00*	0.00*	0.00*
10% betel quid extract	0.02*		0.19	0.00*	0.00*
20% betel quid extract	0.00*	0.19		0.33	0.00*
Positive control	0.00*	0.00*	0.33		0.00*
Negative control	0.00*	0.00*	0.00*	0.00*	

Table 2. The effect of antiseptic of betel quid extract among groups.

*Significant level, Post-hoc Tukey's test, p<0.05.

It could be seen in tabel 2 that all concentration of betel quid extract had significantly difference with negative control in reducing bacterial colonies. It signified that betel quid extracts had antiseptic effect on mucosal wound. Betel quid extract at 20% concentration had no different effect in decreasing the number of bacterial colonies compare to positive control. It was implied that 20% betel quid extract had similar antiseptic effect to 0.2% hyaluronic acid.

Discussion

The existence of antiseptic effect from betel quid extract is due to active ingredients available on betel quid components such as betel leaf, areca nut seed, gambier and mineral slaked lime or betel lime. Betel leaf contains eugenol, essential oil, cavicol, allipyrocatecol and cavibetol compounds that have capability as antiseptic compound.¹⁹ Amalia *et al* found that *Piper betle* infusion was potential as antiseptic due to its phenolic derivatives.²⁰ Eugenol has capability to destroy permeability of bacterial membrane cells resulting in disturbance of bacterial activity and subsequently the death of bacterial cells.^{21,22}

Areca nut seed has antimicrobial effect on several bacteria found in mouth such as *Streptococcus mutans*, *Streptococcus salivarius*, *Candida albicans* and *Fusobacterium nucleatum*.^{23,24} The compound in areca nut seed that is estimated to have antibacterial effect is proantocyanidin, a condensed tannin which belongs to the class of flavonoid.^{24,25} This condensed tannin has ability in inhibiting bacteria found in oral cavity such as *Enterococcus faecalis*, *Phorphyromonas gingivalis* and *Fusobacterium nucleatum*, fungus such as *Candida albicans*, and virus.^{26,27} Tomiyama *et al* revealed that condensed tannin had strong antibacterial actions against bacteria included in biofilms.²⁸

The antiseptic effect of gambier product is due to its chemical compounds, such as catechin and tannin. Dewi *et al* reported that gambier extract had capability to decrease the number of bacterial colonies on teeth which were soaked within artificial saliva.²⁹ Gambier extract takes part in inhibiting gram positive bacteria and less effective in inhibiting gram negative bacteria such as *Escherichia coli*.³⁰ This is due to the fact that cell wall structure of gram negative bacteria is

more complex, because it has three layers consisting of lipoprotein at external layer, polysaccharides at central layer that has a role as barrier for the entrance of antibacterial bioactive compound and peptidoglycan having high lipid content at internal layer. On the other hand, cell wall structure of gram positive bacteria is simpler and consisting of one layer with low lipid content so that catechin can effectively kill gram positive bacteria such as *Streptococcus mutans*.^{31,32} Catechin can inhibit biofilm formation of some bacteria, such as *Streptococcus mutans* and *Lactobacillus*. Catechin binds with protein, damages the cytoplasmic membran, leaks cytoplasmic membran, induces cell lysis and death.³³ Catechin inactivates glucosyltransferase enzymes, blocks the extracellular polysaccharide (EPS) synthesis, reduces the initial formation and accumulation of biofilm, and eliminates the growth of bacteria.^{34,35} Catechin also keeps wound sterile and stimulates the healing process on ulcer.^{35,36,37} Tannin has bacteriostatic and bactericidal against gram positive bacteria, because tannin generates the complexation with the substrate of bacteria. Tannin binds with metal ion involved in the bacteria metabolism, forms chelat and leads toxicity of cells.³⁸

Calcium hydroxide within betel quid has capability to change the environment into alkaline condition. This alkaline condition can affect the environment so that bacteria can not proliferate and produce neutral condition which stimulate the formation of hard tissue due to the decomposition of calcium hydroxide into ions of Ca^{2+} and OH^- .³⁹ Highly alkaline pH was also effective in eliminating resistant bacteria, such as *Enterococcus faecalis*.⁴⁰ The antibacterial mechanisms from calcium hydroxide is directly related to the increase of pH due to the release of ion OH^- . High decomposition of hydroxyde ions kill bacterial cells through damaging of cytoplasm membrane, protein denaturation and DNA damage.⁴¹ Dixit *et al* reported that calcium hydroxide can eliminate bacteria in contaminated area in patients with periapical lesion.⁴²

Conclusion

In conclusion, the study shows that betel quid extract has antiseptic effect on lower lip mucosal wound of male Wistar rats. 20% betel quid extract has equal effect compared to 0.2% hyaluronic acid and it has dose-dependent effect.

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References

1. Junior JCEM, Nunes LHAC, Aruda CS, Rizzi CC, Mouchrek AQS, Tavarez RRDJ, Tonetto MR, Bandeca MC, Filho EMM. Effectiveness of oral antiseptic on tooth biofilm: A study in vivo. *J Contemp Dent Prac.* 2015; 16(8): 674-678.
2. Mummolo S, D'Ercole S, Marchetti E, et al. Oral antiseptic and periodontitis: A clinical and microbiological study. *Oral Health Dent Managemet.* 2014; 13(3): 689-702
3. Roberts CD, Leaper DJ, Assadian O. The role of topical antiseptic agents within antimicrobial stewardship strategies for prevention and treatment of surgical site and chronic open wound infection. *Adv Wound Care.* 2017; 6(2): 63-71
4. Bayoumi AM, Jan A, Al-Amoudi W, Shakir M. The effect of using hyaluronic acid on the extraction sockets. *J Dent Oral Health.* 2015; 2(1):1-5
5. Kapoor P, Sachdeva S, Sachdeva S. Topical hyaluronic acid in the management of oral ulcers. *Indian J Derm.* 2011; 56(3): 300-302
6. Rooney DF. *Betel chewing traditions in South East Asia.* 1st ed. New York: Oxford University Press; 1993: 14-21
7. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. *Betel-quist and areca-nut chewing and some areca-nut derived nitrosamines.* IARC Monogr Eval Carcinog Risks Hum. 2004;85:1-334.
8. Toprani R, Patel D. Betel leaf: Revisiting the benefits of an ancient Indian herbs. *South Asian J Cancer.* 2013; 2(3): 140-141
9. Lutviandhitarani G, Harjanti DW, Wahyono F. Green antibiotics betel leaf (*Piper betle* L.) as a substitute for commercial antibiotics to handle mastitis. *Journal of Agripet.* 2015; 15(1): 28-32
10. Harnowo I, Yunianti. The addition of areca nut seed extract and citric acid on physical, chemical and organoleptical properties of sweet starfruit juice. *J Food Agroind.* 2015; 3(3): 1241-1251
11. Isnawati A, Raini M, Sampurn, Mutiatikum, Widowati L, Gitawati R. Characterization of three types of gambier extract (*Uncaria gambir roxb*) from West Sumatra. *Bulletin Health Res.* 2012; 40(4): 201-208
12. Alphianti L. Apexification treatment using calcium hydroxide paste: 12 months evaluation. *Int Dent J.* 2014; 3(1): 52-59
13. Cwikla SJ, Bellanger M, Giguere S, Fox AP, Vertucci FJ. Dentina tubulus disinfection using three calcium hydroxide formulation. *J Endod.* 2005; 31(1): 50-2
14. Hoque M, Rattila S, Asaduzzaman M, Bari L. Antibacterial activity of ethanol extract of betel leaf (*Piper betle* L.) Against some food borne pathogens. *Bangladesh J Microbiol.* 2011; 28(2): 58-63
15. Rahman MA, Sultana P, Islam MS, Mahmud MT, Rashid MMO, Hossen F. Comparative antimicrobial activity of Areca catechu nut extracts using different extracting solvents. *Bangladesh J Microbiol.* 2014; 31(1): 19-23
16. Katu H, Sumintarti, Mattulada I, Samad R, Hatta M, As'ad S. Inhibitory concentration and minimum contact time gambier extract (*Uncaria gambir Roxb.*) against bacterial growth *Enterococcus faecalis.* *IJSBAR.* 2016; 27(3): 239-246
17. Mohammadi Z, Shalavi S, Yazdizadeh M. Antimicrobial activity of calcium hydroxide in endodontics: a review. *Chonnan Med J.* 2012; 48(3): 133-140
18. Arts JWM, Kramer K, Arndt SS, Ohi F. Sex differences in physiological acclimatization after transfer in Wistar rats. *Animals (Basel).* 2014 Dec; 4(4):693-711
19. Dwidevi V, Tripathi S. Review study on potential activity of *Piper betle.* *J Pharmacog Phytochem.* 2014; 3(4): 93-98
20. Amalia H, Sitompul T, Hutauruk J, Andrianjah, Mun'im A. Effectiveness of *Piper betle* leaf infusion as a palpebral skin antiseptic. *Universa Medicana.* 2009; 28(2): 83-91
21. Nunez L, Aquiono MD. Microbe activity of clove essential oil (*Eugenia caryophyllata*). *Braz J Microbiol.* 2012; 43(4): 1256-1260
22. Devi KP, Nisha SA, Sakthivel R, Pandian SK. Eugenol (an essential oil of clove) acts as an antibacterial agent against *Salmonella typhi* by disrupting the cellular membrane. *J. Ethnopharmacol.* 2010; 130(1): 107-115
23. Lingappa A, Nappalli D, Sujatha GP, Shiva PS. Areca nut: to chew or not to chew. *J Dent.* 2011; 1(3): 46-50
24. Antikhat RRN, Michael A. Study on Areca nut for its antimicrobial properties. *J Young Pharmacist.* 2009; 1(1): 42-45
25. Vanimakhal RR, Ezhilarasi BS. Phytochemical Qualitative Analysis and total tannin content in aqueous extract of Areca catechu nut. *Asian J Biomed Pharmaceutic Sci.* 2016; 6(54): 7-9
26. Djauharie N, Kemala N. Antibacterial efficacy of 5% ethanolic extract of propolis (EEP) solution againsts *Enterococcus faecalis.* *J Int Dent Med Res.* 2017; 10(1): 19-23
27. Sarpangala KB, Sarpangala M, Devasya A. Antimicrobial properties of Areca Nut (*Areca catechu, L*): A Review. *Int J Res Ayurveda Pharm.* 2017; 8(3): 8-12
28. Tomiyama K, Mukai Y, Saito M, Watanabe K, Kumada H, Nihei T, Hamada N, Teranaka T. Antibacterial action of a condensed tannin extracted from astrigent Persimmon as a component of food addictive pansil PS-M on oral polymicrobial biofilm. *Biomed Rev. Int.* 2016; 2016: 5730784
29. Dewi SRP, Kamalludin MT, Theodorus, Pambayun R. Anticariogenic effect of gambier (*Uncaria gambir [Roxb.]*) extract on enamel tooth surface exposed by *Streptococcus mutans.* *Int J Health Sci Res.* 2016; 6(8): 171-179
30. Magdalena NV, Kusnadi J. Antibacterial from Gambier leaves crude extract (*Uncaria gambir var Cubadak*) microwave-assisted extraction method againsts bacterial pathogens. *J Food Agrobusiness.* 2015; 3(1): 124-135
31. Kajiji K, Hojo H, Suzuki M, Nanjo F, Kniazawa S, Nakayama T. Relationship between antibacterial activity of (+)-catechn derivatives and their interaction with a model membrane. *J Agri and Food Chem.* 2004; 52(6): 1514-1519.
32. Pambayun R, Gardjito M, Sudarmadi S, Rahayu K. Sensitivity of gram positive bacteria in catechins extracted from gambier (*Uncaria Gambir [Roxb.]*). *Agritech.* 2008;28(4):174-179.
33. Bai L, Takagi S, Ando T, Yoneyama H, Ito K, Mizugai H, Isogai E. Antimicrobial activity of tea catechin againsts canine oral bacteria and functional mechanisms. *J Vet Med Sci.* 2016; 78(9): 1439-1445
34. Ren Z, Chen L, Li J, Li Y. Inhibition of *Streptococcus mutans* polysaccharides synthesis by molecules targeting glycosyltransferase activity. *J Oral Biol.* 2016; 8(1): 31095
35. Bernal P, Lemaire S, Pinho MG, Mobashery S, Hinds J, Taylor PW. Insertion of epicatechin gallate into the cytoplasmic membrane of methicillin-resistant *Staphylococcus aureus* disrupts penicillin-binding-protein (PBP) 2a-mediated beta lactam resistance by delocalizing PBP2. *J Biol Chem.* 2010;285(31):24055-24065.
36. Arundina I, Diyatri I, Budhy TI, Jit FY. The effect of Brotowali stem extract (*Tinospora crispa*) towards increasing number of lymphocytes in the healing process of traumatic ulcer on diabetic Wistar rats. *J Int Dent Med Res.* 2017; 10(3): 975-980
37. Roestamadji RI, Arundina I, Diyatri I, Sambodo DT, Irmalia WR. Brotowali extract (*Tinospora crispa*) for oral traumatic ulcer in Diabetes Miletus Wistar rats. *J Int Dent Med Res.* 2017; 10(3): 991-996
38. Joseph N, Mirelle AFR, Matchawe C, Patrice DN, Jasophat N. Evaluation of the antimicrobial activity of tannin extracted from barks of *erythrophleum guineensis* (Caesal piniceae). *J Pharmacog Phytochem.* 2016; 5(4): 287-291.

39. Khan S, Inambar NK, Akash Meshram GK, Singh MP, Chaurasia H. Calcium hydroxide-a great calcific wall. J Orofacial Res. 2011; 1(1): 26-30
40. Rusdiana, Usman M, Meidyawati R, Suprastiwi E, Ayu D. Antibacterial effects of bioceramic and mineral trioxide aggregate sealers againsts Enterococcus faecalis clinical isolates. J Int Dent Med Res. 2017; 10(3): 981-986
41. Revathi N, Chandra SSM. Merits and demerits of calcium hydroxide as a therapeutic agents: a review. Int J. Dent. Sci. Res. 2014; 68: 1-4
42. Dixit S, Dixit A, Kumar P. Nonsurgical treatment of two periapical lesion with calcium hydroxide using two different vehicles. Case Reports in Dent. 2014; 2014: 901497.