

Antibacterial Effect of Telang Flower (*Clitoria Ternatea*) Extract in Eradicating *Streptococcus Mutans* UA 159 Biofilm Mass

Jeffrey^{1*}, Rahmadaniah Khaerunnisa², Isti Arifianti³, Nuri Khalish Azhari⁴

1. Department of Pediatric Dentistry, Faculty of Dentistry, Jenderal Achmad Yani University, Cimahi, West Java, Indonesia.
2. Department of Oral Biology, Faculty of Dentistry, Jenderal Achmad Yani University, Cimahi, West Java, Indonesia.
3. Department of Prosthodontics, Faculty of Dentistry, Jenderal Achmad Yani University, Cimahi, West Java, Indonesia.
4. Faculty of Dentistry, Jenderal Achmad Yani University, Cimahi, West Java, Indonesia.

Abstract

Antibacterial activity is indicated by the blue color on the telang flower (*Clitoria ternatea*) because it contains anthocyanins which are included in the flavonoid group. The purpose of this study was to examine the antibacterial activity of telang flower against *Streptococcus mutans* in eradicating *S. mutans* biofilm mass.

This research is a laboratory experimental study on the antibacterial activity of telang flower extract against *Streptococcus mutans* UA 159. The diameter of the inhibition zone was observed and the data obtained from measurement of the percentage of viability and inhibition were then statistically tested using the normality test, homogeneity test, and the Post Hoc Tukey HSD test. Further, calculate the eradication percentage of *Streptococcus mutans* biofilm formation by comparing the Optical Density (OD) value of each treatment.

There is antibacterial activity of the telang flower extract against *S. mutans*, with the value of Minimal Inhibitory Concentration (MIC) and Minimal Bactericidal Concentration (MBC) in this study was at a concentration of 12.5 % and 100 % respectively. Telang flower extract was shown to significantly inhibit the growth of *S. mutans* ($p < 0.05$) and with a concentration of 100% showed biofilm eradication results that were almost close to the positive control group.

Telang flower (*Clitoria ternatea*) extract has an antibacterial effect against and the ability to eradicate *Streptococcus mutans* biofilms. The level of inhibition shown by the telang flower extract on the growth of *S. mutans* was directly proportional to its concentration level, the higher the concentration of the extract, the greater the inhibition produced.

Experimental article (J Int Dent Med Res 2023; 16(2): 628-634)

Keywords: Antibacterial, Biofilm mass, Preventive, *Streptococcus mutans*, Telang flower.

Received date: 25 January 2023

Accept date: 23 February 2023

Introduction

Riset Kesehatan Dasar (RISKESDAS) in 2018 stated that 57.6% of Indonesian people had health problems related to dental and oral health. This can occur due to the lack of attention of the Indonesian people to dental and oral health, causing the incidence of dental and oral diseases to increase. If left untreated, this disease will cause pain, impair function, affect body weight, and disrupt growth and development which will ultimately affect the decrease in quality of life.¹⁻³

Dental and oral diseases are formed due to homeostatic imbalances in the oral cavity, such as changes in microorganisms that are influenced by salivary flow, salivary composition, sugar consumption, fluorine exposure, and also prevention factors. This condition is characterized by an increase in the number of *Streptococcus mutans* which will then colonize the tooth enamel surface.⁴⁻⁸ These bacteria have the ability to synthesize extracellular polysaccharides, produce lactic acid, and form colonies that are firmly attached to the tooth surface, all of which contribute to the formation of biofilms.^{9,10} The ability of *S. mutans* to adhere to tooth surfaces and form biofilms is one of its virulence factors.¹¹ This virulence factor also increases along with its ability to survive in an acidic environment.¹⁰ The virulence factor of *S. mutans* is acid tolerance and production,

*Corresponding author:

Jeffrey
Department of Pediatric Dentistry, Faculty of Dentistry,
Jenderal Achmad Yani University, Cimahi,
West Java, Indonesia
E-mail: jeffrey_dent2000@yahoo.com

attachment of adhesion to cell surface protein, production of glucosyltransferase (GTF), which is involved in the production of glucan, and intracellular polysaccharides. Furthermore, it will cause an increase in dental caries if left untreated due to the formation of a strong and cariogenic biofilm layer.⁴⁻⁸

It is well known that one of the etiological factors of dental caries is a microorganism, namely *Streptococcus mutans* in the oral cavity. One effort in the oral cavity to suppress the growth of bacteria is by using chemical agents in the form of mouthwash, such as chlorhexidine (CHX). Maintenance of dental and oral health can be done chemically by using mouthwash. However, continuous long-term use can irritate the oral mucosa, disrupt the microbial balance in the mouth, unpleasant taste, causing staining of the teeth, and increasing bacterial resistance to antibacterials. As a result, efforts to locate safer, ideal, long-term-useable, and minimally harmful substitute materials remain necessary.^{6,12}

Natural products are a new source of clinically applicable antibacterial agents. The antibacterial agents of plant extracts have been the subject of many reports. Polyphenols, such as phenolic acids, anthocyanins, flavonoids, coumarins, stilbenes, lignans, tannins and lignins, have high antibacterial activity due to their secondary metabolite content. Flowers, fruits, vegetables, and seeds contain a lot of substances that help build resistance to a variety of pathogenic microbes and protect against free radicals and toxins.¹³⁻¹⁵

Plants are a source of pharmacological compounds used as traditional medicine for several diseases such as microbial infections which are a significant public health problem and are always of interest to scientists. Recently many phytochemicals, including antibacterials, have been clarified from edible plants. Several studies have also been reported regarding herbal plant components, which have shown antibacterial activity against *S. mutans*.^{16,17}

Recently, herbal plants are widely used as antibacterials because they have fewer side effects, inexpensive, and effective. One of them is telang flower (*Clitoria ternatea*), a plant that has long been used for treatment because of its active biological content, such as alkaloids, flavonoids, saponins, triterpenoids and steroids which have potential as antibacterials.^{18,19} The

content of flavonoids in this plant is known to have antibacterial properties due to the hydroxyl groups in the structure of flavonoids can cause toxic effects on bacteria because they cause changes in the transportation of organic components and nutrients.^{20,21} Another component is alkaloids which can cause cell death by interfering with peptidoglycan in bacterial cells and preventing the formation of cell wall layers. Triterpenoids can bind to transmembrane proteins, causing damage to the bacterial cell membrane. It is the action of these components that causes the telang flower to be used as an antibacterial agent to prevent the occurrence of infectious diseases.^{22,23}

Regular tooth brushing is one of the mechanical efforts in maintaining oral health which aims to eliminate biofilm but is less effective, therefore an effort is needed to use chemical agents in the form of mouthwash as an effort to prevent dental caries, so as to prevent bacterial growth in the oral cavity. Thus dental and oral health can be maintained by using mouthwash, but its use in the long term can cause side effects, so it is necessary to develop plants as natural ingredients containing biologically active components for oral prophylaxis.^{6,24}

Materials and methods

Laboratory experimental research on the antibacterial of telang flower (*Clitoria ternatea*) extract against *Streptococcus mutans* UA 159 with various concentrations on Mueller Hinton Agar (MHA) media was then carried out by observing the diameter of the inhibition zone and its ability to eradicate *S. mutans* biofilm mass.

The first step is the preparation of growing media using Mueller Hinton Agar (MHA) and Mueller Hinton Broth (MHB). Thirty eight grams of MHA media and 21 grams of MHB media were weighed using an analytical balance. Both were then dissolved respectively in 1000 ml of ddH₂O with the help of a microwave. After that, it was sterilized for 20 minutes in an autoclave at 121°C and 1.5 atm pressure.

Telang flower (*Clitoria ternatea*) was used in this study. Furthermore, the preparation of telang flower extract was carried out by preparing a stock solution, making a series of Working Solution (WS), and sample filtration. The telang flower extract stock solution used was prepared

by dissolving 2 mg of extract paste in 1 ml of 100% DMSO so that the stock solution has a concentration of 2 mg/ml of extract in 100% DMSO.

The Working Solution (WS) series was made by diluting the stock of telang flower extract using ddH₂O then using 10% DMSO to make the concentration series. The extract concentration series used are:

TFE 100%: 50 μ l stock solution + 950 μ l ddH₂O (1st solution)

TFE 50%: 500 μ l 1st solution + 500 μ l DMSO 10% (2nd solution)

TFE 25%: 500 μ l 2nd solution + 500 μ l DMSO 10% (3rd solution)

TFE 12.5%: 500 μ l 3rd solution + 500 μ l DMSO 10% (4th solution)

TFE 6.25%: 500 μ l 4th solution + 500 μ l DMSO 10% (5th solution)

TFE 3.125%: 500 μ l 5th solution + 500 μ l DMSO 10%

The working solution was then filtered using a 0.22 μ m pore filter syringe to obtain a sterile sample.

The direct colony suspension method was used to make *S. mutans* inoculums. Inoculums were obtained by inoculating *S. mutans* colonies on MHA medium into MHB. To obtain an inoculum with a bacterial count of around $1-5 \times 10^8$ CFU/ml, the turbidity of the solution was then adjusted visually to the turbidity of 0.5 McFarland standard solution. Then the solution was diluted using MHB to produce inoculums with bacterial counts in the range 2×10^6 - 1×10^7 CFU/ml with a ratio of 1:50. Then dilution was carried out using MHB media with a ratio of 1:20 to produce an inoculum with a bacterial count of $1-5 \times 10^5$ CFU/ml.

The broth microdilution method was carried out in wells by adding 100 μ l of inoculum of various concentrations of telang flower extract, then adding 100 μ l of the working solution for each concentration of the extract in the well so that the extract concentration reached the final concentration and 100 μ l of chlorhexidine 0.2% was added to the well. Furthermore, for growth control, 100 μ l MHB and 100 μ l inoculum were added to the well. As a blank, add 100 μ l of MHB and 100 μ l of working solution to the well for each concentration of telang extract, then also make a 0.2% Chlorhexidine blank. Incubate the plate for 24 hours in an incubator at 37°C, followed by measurements using

spectrophotometry with a wavelength of 530 nm. The Optical Density (OD) values of the treatments and each blank were compared to determine the growth of *S. mutans*. The lowest extract concentration with a 99 percent inhibition of bacterial growth was used to calculate the MIC value.

The pour plate method was carried out after spectrophotometric measurements of each treatment in the microplate. This method is carried out to prove that the minimum concentration of bactericidal observed in spectrophotometric measurements is able to provide an inhibitory effect so that there is no growth of bacterial colonies on the agar medium. Then carry out the pour plate method by taking 20 μ l of solution from each well and growing it on MHA media, then incubating at 37°C for 24 hours. Visual observations were made then the number of colonies of bacteria was counted using a colony counter. The MIC value was proven by the concentration of the extract which was able to inhibit bacterial growth so that no growth of bacterial colonies was observed on the agar medium.

The data obtained from the measurement of the percentage of viability and inhibition were statistically tested. In this study, the statistical tests used were the normality test, homogeneity test, and the Post Hoc Tukey HSD test. The IBM Statistics SPSS 25 application is used to perform statistical tests.

The Streptococcus mutans biofilm eradication test was then carried out by culturing Streptococcus mutans on Brain Heart Infusion Medium at 37°C for 24 hours until the bacteria were at 0.5 Mc Farland turbidity. As much as 50 μ L of Streptococcus mutans was put into 96 well plates then added 100 μ L of telang flower extract with each concentration into 96 well plates which already contained the bacterial culture. Incubation for 48 hours at 37°C. After incubation was complete, the microplate incubation liquid was removed and then washed with PBS solution 2 times. After that, 125 μ L of Crystal Violet 0.1% was added to the well and incubated for 10-15 minutes, the microplate was rinsed using distilled water 3-4 times. Then 125 μ L of 30% acetic acid was added to the well plate which had been stained with 0.1% Crystal Violet. Then measured with a microplate reader with a wavelength of 490 nm. By comparing the OD values of each treatment and each blank, biofilm growth was

identified.

The formula used to calculate the eradication percentage for Streptococcus mutans biofilm formation is:

$$\% \text{ inhibition} = \left[1 - \frac{OD \text{ sample} - OD \text{ sample blank}}{(OD \text{ solvent} - OD \text{ solvent blank})} \right] \times 100\%$$

Information:

OD sample : Optical density of telang flower extract + solvent + bacterial suspension.

OD sample blank: Optical density of telang flower extract + solvent.

OD solvent : Optical density control of solvent + bacterial suspension.

OD solvent blank: Optical density control of solvent.

The examination was carried out 3 times and the average OD value was determined. Biofilm formation is measured by:
 Biofilm formation = OD at 490nm in bacterial assay – OD control

Results

The results of this study indicate that there is an antibacterial activity of telang flower (*C. ternatea*) extract against *S. mutans*. In this study, the MIC and MBC of telang flower extract against *S. mutans* was 12.5% and 100%, respectively, as can be seen in Table 1. The value of the MBC of telang flower extract against *S. mutans* in this study was 100% as shown in Table 1.

Sample	Viability (%)	Inhibition (%)
TFE 100%	1.35 ± 0.10 ^a	98.65 ± 0.10 ^a
TFE 50%	8.44 ± 0.36 ^b	91.56 ± 0.36 ^f
TFE 25%	15.75 ± 0.43 ^c	84.25 ± 0.43 ^e
TFE 12.5%	41.67 ± 0.60 ^d	58.33 ± 0.60 ^d
TFE 6.25%	90.47 ± 0.59 ^e	9.53 ± 0.59 ^c
TFE 3.125%	97.47 ± 0.24 ^f	2.53 ± 0.24 ^b
GC (Growth Control)	100.00 ± 0.19 ^g	0.00 ± 0.19 ^a
Chlorhexidine 0.2%	1.25 ± 0.19 ^a	98.75 ± 0.19 ^g

Table 1. Viability and Inhibition of Telang Flower Extract (TFE) against *S. mutans*.

*Data presented is the average ± standard deviation. Letters (a,b,c,d,e,f,g) indicate a significant difference based on the Tukey HSD test (p<0.05).

The extract of the telang flower (*C. ternatea*) was shown to inhibit the growth of *S. mutans* significantly (p<0.05) as shown in Figures 1 and 2. The inhibition ability of telang flower extract on the growth of *S. mutans* bacteria can be said to be directly proportional to its concentration, the inhibitory effect will be

higher if the concentration of the extract is also higher.

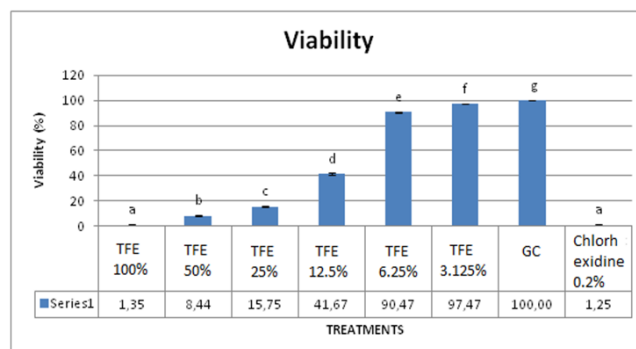


Figure 1. Graph of Viability Percentage of *S. mutans* after administration of Telang Flower Extract.

*Data presented is the average ± standard deviation. Letters (a,b,c,d,e,f,g) indicate a significant difference based on the Tukey HSD test (p<0.05).

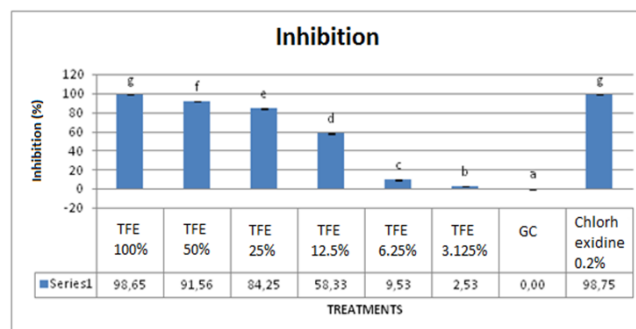


Figure 2. Percentage of Inhibition of Telang Flower Extract against *S. mutans*.

*Data presented is the average ± standard deviation. Letters (a,b,c,d,e,f,g) indicate a significant difference based on the Tukey HSD test (p<0.05).

Sample	Dilution factor	Number of colonies			CFU/ml			Average	Average
		1	2	3	1	2	3		
PC	10000	0	0	0	0	0	0	0	
GC	10000	TNTC	TNTC	TNTC	TNTC	TNTC	TNTC	TNTC	
TFE 100%	10000	3	0	0	3 x 10 ⁴	0	1.00	1 x 10 ⁴	
TFE 50%	10000	25	20	29	25 x 10 ⁴	20 x 10 ⁴	29 x 10 ⁴	24.67	
TFE 25%	10000	40	35	37	40 x 10 ⁴	35 x 10 ⁴	37 x 10 ⁴	37.33	
TFE 12.5%	10000	79	78	82	79 x 10 ⁴	78 x 10 ⁴	82 x 10 ⁴	79.67	
TFE 6.125%	10000	147	151	149	147 x 10 ⁴	151 x 10 ⁴	149 x 10 ⁴	149.00	
TFE 3.125%	10000	190	198	199	190 x 10 ⁴	198 x 10 ⁴	199 x 10 ⁴	195.67	

Table 2. Calculation of the number of colonies of telang flower extract in the MIC and MBC tests.

TNTC : Too Numerous To Count

PC (Positif Control): *S. mutans* inoculum + Chlorhexidine 0.2%

GC (Growth Control): *S. mutans* inoculum + MHB (Mueller Hinton Broth)

Streptococcus mutans biofilm eradication testing showed that all treatment groups were able to eradicate biofilms with the highest inhibition percentage being the telang flower extract treatment group with a concentration of 100%. While the treatment group that showed

the lowest results in eradicating *S. mutans* biofilm was the telang flower extract treatment group with a concentration of 3.125%. (Table 3)

Group	Inhibition (%)			Average
	1	2	3	
Negative control	3.36	3.33	3.28	3.33
TFE 3.125%	4.27	4.18	4.10	4.19
TFE 6.25%	12.26	12.25	12.16	12.23
TFE 12.5%	53.75	53.72	52.46	53.31
TFE 25%	88.02	87.92	87.93	87.96
TFE 50%	94.64	94.60	94.59	94.61
TFE 100%	99.96	99.98	99.93	99.96
Positive control	100.00	100.00	100.00	100.00

Table 3. Results of Biofilm Eradication Test of Telang Flower Extract.

Discussion

The *Clitoria ternatea* plant is widely used as a traditional herbal medicine in Indonesia. Although all parts of the plant can be used, the flower parts are most often used due to their antimicrobial, anti-inflammatory, and antioxidant properties. The antibacterial potential of various parts of the *C. ternatea* plant in maintaining oral health is not known with certainty, therefore this study was conducted to determine the antibacterial activity of telang flower extract on the growth of *Streptococcus mutans* bacteria because this bacterium plays a very important role in the occurrence of dental caries.⁵⁻⁸

Herbal plants such as telang flower used alone or in combination have been scientifically proven to be safe and effective drugs without side effects. The main advantage of using herbal plants is the minimal side effects and so far there have been no reports of any side effects.²⁵ Telang flower extract has an antibacterial activity which results in a decrease in the number of bacteria so that the ability of bacteria to colonize and communicate with each other is hampered.^{26,27}

Dental caries is thought to be caused by *Streptococcus mutans*, which can form biofilms on the surface of teeth. Its capacity to obtain nutrients and metabolize fermentable dietary carbohydrates into acids contributes to its pathogenicity. The response of *S. mutans* to environmental stress is critical for its virulence and survival. The VicRK system is one of the two-component signal transduction systems (TCS) that is thought to exist in *S. mutans*. The function of the VicRK signal transduction system

is the main regulatory factor for acidification, cell wall metabolism, biofilm formation, and bacterial oxidative stress response so that it can cause *S. mutans* to survive.^{28,29}

Chlorhexidine is a gold standard antimicrobial that acts as a very broad spectrum antimicrobial, considered as an agent for evaluating the effectiveness of other antimicrobials, antiplaques, and herbal extracts, so this study was used as a positive control. Chlorhexidine is considered one of the best antimicrobial agents in controlling the growth of oral bacteria.³⁰⁻³⁵ Chlorhexidine, like other drugs, is certainly not without side effects. Chlorhexidine and herbs are both effective for biofilm control in preventing plaque and gingivitis. However, due to the reported side effects due to the use of chlorhexidine as well as the good biocompatibility and acceptability of herbs, herbs can be used effectively as alternatives to chlorhexidine.³⁶

In this study, the percentage inhibition of *S. mutans* flower extract against *S. mutans* at a concentration of 100% was as good as 0.2% Chlorhexidine (Tables 1 and 2). Based on table 3, telang flower extract can eradicate *Streptococcus mutans* biofilms. The telang extract treatment group with a concentration of 100% showed the highest results among other concentrations with an average of 99.96% and almost close to the positive control group which had an average value of 100.00%. While the telang extract treatment group with a concentration of 3.125% showed the lowest results compared to other concentrations, namely an average of 4.19%. This is because the highest content of telang flower extract is flavonoids, saponins, terpenoids, and alkaloids.^{19,21,31,37-39}

Flavonoids can inhibit the function of cell membranes and penetrate the bacterial lipid bilayer, thus damaging the outer membrane barrier function. Furthermore, these compounds cause membrane fusion and result in cell leakage. However, this compound was less successful at penetrating the lipopolysaccharide membrane of gram-negative bacteria. Therefore, the antibacterial mechanism is limited. In another study, flavonoids inhibited bacterial energy metabolism which is necessary for the synthesis of macromolecules (DNA, RNA, and proteins).^{20,31}

The role of saponins as an antibacterial is by denaturing proteins by reducing the surface tension of the bacterial cell wall which causes

lysis, damage or disturbance to the permeability of the bacterial cell membrane.^{40,41} In addition to saponins, terpenoids also have antibacterial activity by destroying transmembrane proteins through their reactions with transmembrane proteins on the outer membrane of bacterial cells and forming strong polymeric bonds.^{42,43}

Terpenoids are carbon dioxide, meaning they contain the elements carbon and hydrogen which have been denatured by oxidation or chemically modified. Terpenoids represent a large class of antibacterial compounds produced by plants. Terpenoids originate from five-carbon isoprene units and most of the terpenoids have multicyclic structures that differ from one another in their functional groups and their basic carbon skeleton.⁴²⁻⁴⁴

The structure of other compounds, namely alkaloids which are derivatives of furoquinoline, these compounds can cause interference with the formation of the bacterial cell wall layer by interfering with the constituent components of peptidoglycan will cause cell death and depolarizing the cell wall.^{22,45,46} In this plant tannins have phenol groups, these components can form complex compounds, which cause bacterial cells to die due to the incomplete formation of bacterial cell walls.^{47,48}

The ability of telang flower extract to react with cell membrane activity, energy production, the synthesis of structural components, and disrupting the function of genetic material, can cause disruption of the activity of the glucosyltransferase enzyme used by *S. mutans* to synthesize sucrose in glucan medium which is a bacterial attachment medium. If the amount of glucan is small, it will inhibit the formation of biofilms.⁴⁹⁻⁵¹

Conclusions

Telang flower (*Clitoria ternatea*) extract has an antibacterial effect against and the ability to eradicate *Streptococcus mutans* biofilms. The level of inhibition shown by the telang flower extract on the growth of *S. mutans* was directly proportional to its concentration level, the higher the concentration of the extract, the greater the inhibition produced.

Acknowledgments

The authors would like to thank the

Faculty of Dentistry, Jenderal Achmad Yani University.

Declaration of Interest

The authors declare no conflict of interest.

References

1. Indonesian Ministry of Health. Basic Health Research (RISKESDAS 2018). ed. 2018:93-94.
2. Ozdemir D. Dental Caries and Preventive Strategies. *J Educ Instr Stud*. 2014;4(4):20-24.
3. Rizwan Ullah MSZ. Oral and Dental Delivery of Fluoride: A Review. *Res Rev Fluoride*. 2015;48(3):195-204.
4. Arévalo-Ruano ML, Melo FYC, Echeverry-Chica J, Salazar CL, Martinez-Delgado CM, Pabon MCM, et al. Molecular Identification and Genotyping of *Streptococcus mutans* from Saliva Samples of Children in Medellin, Colombia. *Rev CES Odontol*. 2014;27(2):47-60.
5. David J. Bradshaw. Diet and The Microbial Aetiology of Dental Caries: New Paradigms. *Int Dent J*. 2013;63(s2):64-72.
6. Lee YH, Chung SW, Auh Q, Hong SJ, Lee YA, Jung J, et al. Progress in Oral Microbiome Related to Oral and Systemic Diseases: An Update. *Diagnostics*. 2021; 11(7):1283.
7. Bowen. Biology of *Streptococcus mutans*-Derived Glucosyltransferases: Role in Extracellular Matrix Formation of Cariogenic Biofilms. *Caries Res*. 2011;45(1):69-86.
8. Jeffrey J, Satari M, and Kurnia D, Sudigdoadi S. Inhibition of *Streptococcus mutans* Growth Induced by the Extract of *Citrus aurantifolia* peel. *JIDMR*. 2020;13(1):122-7.
9. Jeffrey, Satari MH, and Kurnia D. Antibacterial effect of lime (*Citrus aurantifolia*) peel extract in preventing biofilm formation. *J Med Heal*. 2019;2(4):1020-9.
10. Sionov RV, Tsavdaridou D, Aqawi M, Zaks B, Steinberg D, and Shalish M. Tooth mousse containing casein phosphopeptide-amorphous calcium phosphate prevents biofilm formation of *Streptococcus mutans*. *BMC Oral Health*. 2021;21(1):1-10.
11. Kunarti S, Ramadhani A, and Setyowati L. Antibiofilm activity of mangosteen (*Garcinia mangostana* L.) flavonoids against *Streptococcus mutans* bacteria. *Conserv Dent J*. 2020;10(2):48-50.
12. Neves O.V.M, Suwindere W, and Sugiaman V.K. The Differences in Antibacterial Activity of Ethanol Extracts of Green Betel Leaves (*Piper betle* L.) and Lemongrass Leaves (*Cymbopogon citratus*) Against *Streptococcus mutans* In Vitro. *Cakradonya Dent J*. 2012;14(2):69-76.
13. Livia Slobodníková, Katarína Rendeková, Ján Kováč, and Pavel Mučaji. Antibiofilm Activity of Plant Polyphenols. *Molecules*. 2016;21(12):1717. DOI: 10.3390/molecules21121717.
14. Caballero, Paul BF. *Encyclopedia of Food and Health*. Elsevier Oxford; 2016:40.
15. Stan D, Enciu AM, Mateescu AL, Cristina Ion A, Brezeanu AC, Cristina A, et al. Natural Compounds With Antimicrobial and Antiviral Effect and Nanocarriers Used for Their Transportation. *Frontiers in Pharmacology*. 2021;12:1-25.
16. Mohamed SG. Antimicrobial Activity of *Syzygium aromaticum* and *Citrus aurantifolia* Essential Oils Against Some Microbes in Khartoum, Sudan. *EC Microbiology*. 2017;6:253-259.
17. Marsh PD, Lewis MAO, Rogers H, Williams DW, and Wilson M. *Marsh and Martin's Oral Microbiology*. 6th ed. Elsevier. 2016:112-158.
18. Suwandi T. The initial treatment of mobile teeth closure diastema in chronic adult periodontitis. *Jurnal PDGI*. 2010;59(3):105-109. <https://www.academia.edu/download/40707563/12-44-1-PB.pdf>
19. Jeyaraj EJ, Lim YY, and Choo WS. Extraction methods of telang (*Clitoria ternatea*) flower and biological activities of its phytochemicals. *J Food Sci Technol*. 2021;58(6):2054-67.

- <https://doi.org/10.1007/s13197-020-04745-3>.
20. Xie Y, Yang W, Tang F, Chen X, and Ren L. Antibacterial activities of flavonoids: structure-activity relationship and mechanism. *Current Medicinal Chemistry*. 2015;22(1):132-149.
 21. Jeyaraj EJ, Nathan S, Lim YY, and Choo WS. Antibiofilm properties of *Clitoria ternatea* flower anthocyanin-rich fraction towards *Pseudomonas aeruginosa*. *Access Microbiology* 2022;4:000343. DOI 10.1099/acmi.0.000343.
 22. Amajida H, Purwoko T, and Susilowati A. Antibacterial activity of ethanolic and n-hexane extracts of *Ruellia tuberosa* leaves against *Escherichia coli* and *Bacillus subtilis* bacteria. *Biofarmasi J Nat Prod Biochem*. 2019;17(2):69-80.
 23. Dacullo MRA and Bitacura JG. In Vitro Antibacterial, Anticoagulant, and Antioxidant Screening of Aqueous Extracts of Blue Ternate (*Clitoria ternatea* L.) Flower. *Herbal Medicines Journal*. 2022;7(4):1-10. DOI: <https://doi.org/10.22087/hmj.v7i3.867>
 24. Attamimi FA, Ruslami R, and Maskoen AM. Antibacterial Activity Test of Ant Nest Tuber (*Myrmecodia Pendens*) Crude Extract Against *Streptococcus sanguinis* Compared to Chlorhexidine. *Bandung Medical Journal*. 2015;49(2):94-101.
 25. Rafiq S, Kaul R, Sofi SA, Bashir N, Nazir F, and Ahmad G. Citrus peel as a source of functional ingredient: A review. *J Saudi Soc Agric Sci*. 2018;17(4):351-358. doi:10.1016/j.jssas.2016.07.006
 26. Anitha L. and Sunitaraju K. A Review on Antimicrobial Activity of Vegetables, Herbs and Spices Against Cariogenic Bacteria. *Research & Reviews: Journal of Biology*. 2016;4(4):12-20.
 27. Boakye YD, Osafo N, Danquah CA, Adu F, and Agyare C. Antimicrobial Agents: Antibacterial Agents, Anti-biofilm Agents, Antibacterial Natural Compounds, and Antibacterial Chemicals. *Antimicrobial, Antibiotic Resistance, Antibiofilm Strategies and Activity Methods*. 2019:1-24. Editor: Kirmusaoğlu S. doi:10.5772/intechopen.82560.
 28. Lei L, Long L, Xin Yang, Yang Qiu, Yanglin Z, Tao Hu, et al. The VicRK Two-Component System Regulates *Streptococcus mutans* Virulence. *Curr. Issues Mol. Biol*. 2019;32:167-200. doi: <https://doi.org/10.21775/cimb.032.167>
 29. Alves L, Erika N, Thais H, Rafael N, Flávia S., José F, et al. The two-component system VicRK regulates functions associated with *Streptococcus mutans* resistance to complement immunity. *Mol Oral Microbiol*. 2017;32(5):419-431. doi:10.1111/omi.12183
 30. Rekha SR, Kulanthaivel M, and Hridhya KV. Antibacterial Efficacy And Minimum Inhibitory Concentrations of Medicinal Plants against Wound Pathogens. *Biomed. & Pharmacol. J*. 2018;11(1):237-246. <http://dx.doi.org/10.13005/bpj/1368>.
 31. Haditio SM, Muttaqin Z, and Hadi L. Comparison of Inhibition Zones Between Telang Flower (*Clitoria ternatea*) and Lemongrass (*Cymbopogon citratus*) Against *Streptococcus mutans* and *Staphylococcus aureus*. *Biomedical Journal of Indonesia*. 2021; 7(2): 374-8. <https://doi.org/10.32539/BJI.v7i2.313>.
 32. Rashid S, Tong WY, Leong CR, Ghazali NM, Taher A, Ahmad N, Tan WN, Teo SH. Anthocyanin Microcapsule from *Clitoria ternatea*: Potential Bio-preservative and Blue Colorant for Baked Food Products. *Arabian Journal for Science and Engineering*. 2020;46:65-72. <https://doi.org/10.1007/s13369-020-04716-y>.
 33. Rafiq S, Kaul R, Sofi SA, Bashir N, Nazir F, Ahmad G. Citrus peel as a source of functional ingredient: A review. *J Saudi Soc Agric Sci*. 2018;17(4):351-358. doi:10.1016/j.jssas.2016.07.006
 34. Mathur S, Mathur T, Srivastava R, Khatri R. Chlorhexidine: The Gold Standard in Chemical Plaque Control. *National Journal of Physiology, Pharmacy & Pharmacology*. 2011;1(2):45-50.
 35. Prasad K, John S, Deepika V, Dwijendra KS, Reddy BR, Chincholi S. Anti-Plaque Efficacy of Herbal and 0.2% Chlorhexidine Gluconate Mouthwash: A Comparative Study. *J Int Oral Health*. 2015;7(8):98-102. PMID: 26464549; PMCID: PMC4588801.
 36. Abouassi T, Hannig C, Mahncke K, et al. Does human saliva decrease the antimicrobial activity of chlorhexidine against oral bacteria. *BMC Research Notes*. 2014;7:1-6.
 37. Oguis GK, Gilding EK, Jackson MA and Craik DJ. Telang (*Clitoria ternatea*), a Cyclotide-Bearing Plant with Applications in Agriculture and Medicine. *Front. Plant Sci*. 2019;10:645. doi: 10.3389/fpls.2019.00645.
 38. Indrianiingsih AW, Wulanjati MP, Windarsih A, Bhattacharjya DK, Suzuki T, Katayama T. In vitro studies of antioxidant, antidiabetic, and antibacterial activities of *Theobroma cacao*, *Annona muricata* and *Clitoria ternatea*. *Biocatalysis and Agricultural Biotechnology*. 2021;33:1-8. <https://doi.org/10.1016/j.bcab.2021.101995>.
 39. Satria D., Sofyanti E., Wulandari P., Fajarini, Pakpahan S.D., and Limbong S.A. Antibacterial activity of Medan Telang (*Clitoria ternatea* L.) corolla extract against *Streptococcus mutans* ATCC®25175™ and *Staphylococcus aureus* ATCC®6538™. *Pharmacia*.2022; 69(1): 195–202. DOI 10.3897/pharmacia.69.e77076.
 40. Khan MJ, Ahmmed A, Shin JH, Baek JS, Kim MY, and Kim JD. Green Tea Seed Isolated Saponins Exerts Antibacterial Effects against Various Strains of Gram Positive and Gram Negative Bacteria, a Comprehensive Study *In Vitro* and *In Vivo*. *Evid Based Complement Alternat Med*. 2018: 1-12.
 41. Nugraha SE, Achmad S, and Sitompul E. Antibacterial Activity of Ethyl Acetate Fraction of Passion Fruit Peel (*Passiflora edulis* Sims) on *Staphylococcus Aureus* and *Escherichia Coli*. *Indonesian Journal of Pharmaceutical and Clinical Research (IDJPCR)*. 2019; 02(1): 07–12.
 42. Nassar, Zeyad, and Abdalrahim. The Pharmacological Properties of terpenoid from *Sandoricum Koetjape*. *J Medcentral*. 2010;1(2):1-11.
 43. Apridamayanti P, Sari R, Rachmaningtyas A, and Aranthi V. Antioxidant, antibacterial activity, and FICI (Fractional Inhibitory Concentration Index) of ethanolic extract of *Melastoma malabathricum* leaves with amoxicillin against pathogenic bacteria. *Nusantara Bioscience*. 2021; 13(2): 140-147.
 44. Barbieri R, Coppo E, Marchese A, et al. Phytochemicals for human disease: An update on plant-derived compounds antibacterial activity. *Microbiol Res*. 2017;196:44-68. doi:10.1016/j.micres.2016.12.003
 45. Jafaar HJ, Isbilen O, Volkan E & Sariyar G. Alkaloid profiling and antimicrobial activities of *Papaver glaucum* and *P. Decaisnei*. *BMC Res Notes*. 2021; 14: 348.
 46. Sulaiman M, Jannat K, Nissapatorn V, Rahmatullah M, Paul AK, Pereira ML, et al. Antibacterial and Antifungal Alkaloids from Asian Angiosperms: Distribution, Mechanisms of Action, Structure-Activity, and Clinical Potentials. *Antibiotics*. 2022; 11: 1146.
 47. Nowak A, Wasilkowski D, and Mrozik A. Implications of Bacterial Adaptation to Phenol Degradation under Suboptimal Culture Conditions Involving *Stenotrophomonas maltophilia* KB2 and *Pseudomonas moorei* KB4. *Water* 2022, 14, 2845. <https://doi.org/10.3390/w14182845>.
 48. Utomo S.B. Antibacterial Activity Test of the C-4-methoxyphenylcalix[4]resorcinarene Compound Modified by Hexadecyltrimethylammonium-Bromide against *Staphylococcus aureus* and *Escherichia coli* Bacteria. *J UNS*. 2018;1(2):77.
 49. Slobodniková L, Fialová S, Rendeková K, Kovář J. Antibiofilm Activity of Plant Polyphenols. *Molecules*. 2016:1-15. doi:10.3390/molecules21121717
 50. Alibi S, Crespo D, Navas J. Plant-derivatives small molecules with antibacterial activity. *Antibiotics*. 2021;10(3):1-19. doi:10.3390/antibiotics10030231.
 51. Elsayed M.A., Elbanna A., Rahman MM, and Elmesellawy M.Y. Comparative Evaluation of the Antibacterial Effect of Different Combinations of Etidronate, Nanochitosan and NaOCl on *E. Faecalis* Biofilm. *JIDMR*. 2022;15(4):1429-33.